Use of seaweed-based biostimulant (*Ascophyllum nodosum*) on ornamental sunflower seed germination and seedling growth

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**Abstract**

Seaweed extracts are employed as biostimulants due to their beneficial effects on crop growth and yield. *Ascophyllum nodosum* seaweed extract aid to improve seedling growth and development, and decrease seedlings production costs; however, the correct concentration must be used in order to maximize the biostimulant effects. Consequently, this study aimed to analyze the effects of different concentrations of a seaweed-based (*Ascophyllum nodosum*) biostimulant on ornamental sunflower seed germination and seedling growth. Seeds of ornamental sunflower cv. “Sol Pleno” were sown in polyethylene trays containing commercial substrate. The treatments consisted of dairy spraying 60 mL of the solutions 0 (control), 5, 10 or 15 mL L⁻¹ biostimulant on substrate. The experimental design was completely randomized with 4 treatments (concentrations of biostimulant) and 4 replicates (10 seeds replicate⁻¹). The evaluated variables were percentage, index and time averages of germination, seedling height, fresh and dry mass of shoot and roots, and root system morphology (WinRhizo). The increase of the biostimulant concentration enhances seed germination and seedlings development. The concentration 15 mL L⁻¹ biostimulant showed the best results for percentage and index of germination and the lowest mean germination time and increase plant height and fresh and dry mass of shoots in relation to the control treatment. Accordingly, 15 mL L⁻¹ biostimulant (*Ascophyllum nodosum*) is recommended for ornamental sunflower “Sol Pleno” seed germination and seedlings growth.

**Keywords**: Helianthus annuus, seeding production, seaweed extract, seed germination.
Introduction

Brazil is among the 15 principal producers of ornamental plants in the world with more than 8 thousand producers cultivating around 350 species and 3 thousand varieties (Schoenmaker, 2019). Floriculture industry is particularly concentrated in the Southeast region, with 45% of the production, 28% of the producers and 21% of the cultivated area in São Paulo State (SEBRAE, 2015; Neves and Pinto, 2015). The ornamental plants production has a high cost-effectiveness if compared to other horticultural crops (Gomes, 2006).

Sunflower is an important industrial crop for oil production, although in recent years it is seeing an increase in its use as a cut flower and dwarf cultivars as pot plant (Cormenzana, 2001). Commercial cut flower sunflowers occupy 400 ha in the UK, with a farmgate value estimate over £7 millions in 2016 (Hanks, 2017). Currently, the average cost of a standard sunflower in EUA is US$7.50 for five stems wholesale (U.S. Department of Agriculture, 2017). In Brazil, sunflower and other so-called field flowers that resemble the sunflower are most widely purchased for Easter, being the symbol flower of this date (Junqueira and Peetz, 2017).

The production of seedlings of ornamental plants directly impacts crop growth and yield (Reis et al., 2016). Seed propagation used in the commercial production of ornamental plants guarantees genetic diversity, low cost and easy transport (Grolli, 2008). One of the challenges of the floricultural market is to increase crop productivity by reducing costs without compromising product quality. New approaches have been proposed to stimulate the sustainability of agricultural products and improve the quality of crops and crop-derived products (Ertani et al., 2018). In this sense, alternatives are required in order to increase plant production through new technologies, such as the use of seaweed-based biostimulants.

The use of biostimulants may enhance crop production by increasing the germination rate and the physiological quality and uniformity of plants. Biostimulants also plays an important role in the early stages of plant development, increasing root growth and plant resistance to environmental stresses (Lana et al., 2009). Biostimulant substances from microorganisms are available in the market, and biostimulant efficiency depends of the target species, concentration applied and formulation of the product used (Du Jardin, 2015).

Seaweed has been used as fertilizers or biostimulants in several crops (Sangha et al., 2014). They are composed of macro and microelements, phytohormones such as auxins, cytokines or gibberellins, which can stimulate plant growth efficiently (Górka, 2018). *Ascophyllum nodosum* is a brown algae collected from the east to the south coast of Canada in an area of approximately 4,000 km (Ugarte et al., 2010) and its extracts produce an array of positive responses in plant physiological processes, such as improving nutrient uptake, root development and photosynthetic activity, as well as producing bioactive molecules acting to prevent plant diseases (Talamini and Stadnik, 2004). Thus, this study aimed to analyze the effects of different concentrations of seaweed-based (*Ascophyllum nodosum*) biostimulant on the seed germination and seedling growth of ornamental sunflower cv. “Pleno Sol”.

Material and Methods

The seeds of ornamental sunflower “Pleno Sol” were purchased from a commercial producer and sown in polyethylene plastic trays (128 cells) filled with commercial substrate (Table 1).

### Material and Methods

The seaweed-based (*Ascophyllum nodosum*) biostimulant was used for experimentation on concentrations of 0, 5, 10 or 15 mL L⁻¹ biostimulant diluted on tap water (Carvalho et al., 2018). The experiment was daily sprayed with 60 mL of the experimental solutions. This practice was carried out during 23 days, after which the evaluations of seed germination and seedling growth were carried out:

- Percentage of germination (%) - calculated by the formula \( G = (N/40) \times 100 \), where: \( N \) = number of seeds germinated at the end of evaluation (germinated seedlings with height at least 2 mm);
- Germination speed index (GSI) - calculated by formula \( \text{GSI} = \sum (n_i/t_i) \), where: \( n_i \) = number of seeds germinating at date ‘i’; \( t_i \) = date after test installation (Maguirre, 1962);
- Mean germination time (TMG, days) - calculated by the formula \( \text{TMG} = (\bar{n}/n_i) / (\bar{t}_i) \), where: \( \bar{n} = \text{mean number of seeds germinated per day}; \bar{t}_i = \text{time after test installation}; n = \text{total number of germinated seeds} \) (Labouriau, 1983).

The analyzed variables were seedling height, fresh and dry (forced-air oven dryer at 65 °C until constant weight) mass of shoots and roots. The analysis of root system morphology was determined by length and the diameter of the root with an optical scanner reader (WinRhizo image analysis system).

The treatments were a completely randomized design with 4 treatments (seaweed-based stimulant) and 4 replicates (10 seeds replicate). Data were subjected to...
Results and Discussion

The best results for all seed germination variables (Table 2) were obtained by the highest biostimulant concentration (15 mL L⁻¹), with 300% higher percentage of germination, 500% higher germination speed index (GSI), and shortened the mean time of germination (MTG) by half in relation to the control treatment. However, all results were below the ornamental sunflower germination data observed by Santos and Castilho (2018) from 92.18% to 96.87%, and Silva et al. (2017) from 75% to 91%. The concentration 15 mL L⁻¹ biostimulant was the only treatment that resulted in more than 50% of germinated seeds; thus, increasing seedling production. Similar effects to the present study were observed in other crops by Gehling et al. (2017) in soybean (Glycine max) and Gehling et al. (2014) in wheat (Triticum aestivum), as the authors verified that higher concentrations of A. nodosum seaweed extract improved seed germination.

Table 2. Percentage of germination (PG), germination speed index (GSI), mean time of germination (MTG) of ornamental sunflower seeds in relation to biostimulants doses.

<table>
<thead>
<tr>
<th>Treatment (ml L⁻¹ biostimulant)</th>
<th>PG</th>
<th>IVG</th>
<th>MTG</th>
</tr>
</thead>
<tbody>
<tr>
<td>0</td>
<td>20.00 b</td>
<td>0.21 b</td>
<td>10.0 a</td>
</tr>
<tr>
<td>5</td>
<td>22.50 b</td>
<td>0.22 b</td>
<td>10.4 a</td>
</tr>
<tr>
<td>10</td>
<td>25.00 b</td>
<td>0.31 b</td>
<td>8.3 a</td>
</tr>
<tr>
<td>15</td>
<td>60.00 a</td>
<td>1.13 a</td>
<td>5.6 b</td>
</tr>
<tr>
<td>DMS</td>
<td>18.93</td>
<td>0.39</td>
<td>2.2</td>
</tr>
<tr>
<td>CV (%)</td>
<td>28.28</td>
<td>39.78</td>
<td>12.1</td>
</tr>
<tr>
<td>F</td>
<td>17.51**</td>
<td>22.57**</td>
<td>17.39**</td>
</tr>
</tbody>
</table>

Means followed by the same do not differ among themselves at the 5% level of significance by the Tukey test. ** - significant at 1%.

The germination rate and mean germination time are inversely proportional; thus, higher the GSI means lower TMG (Santos et al., 2019); accordingly, seeds treated with 15 mL L⁻¹ biostimulant germinated earlier (5.6 days) and higher speed (1.13). Seeds with high GSI and low TMG values are more resistant to the adverse germination conditions, as they germinate faster and the initial stage of development of seedling production is decreased (Oliveira et al., 2009). The GSI and TMG germination data for 15 mL L⁻¹ bioestimulant are very similar to the best results observed by Cabral and Castilho (2016) on ornamental sunflower germination (1.10 of GSI and 89% of germination). Seaweed extracts have the beneficial effect to improve seed germination and seedling establishment making plant propagation more efficient (Sharma et al., 2014). A seedling production program requires a high germination rate in a shorter time (Paim et al., 2016), and the extract compounds increase respiratory rate in seeds, and consequently, the velocity of seedling emergence (Rayorath et al., 2008a). The concentration 15 mL L⁻¹ biostimulant was the first treatment to germinate, initiating seed emergency 4 days after sown. Positions 1 and 2 had the lowest results with seed germination at 12th days (Figure 1).
USE OF SEAWEED-BASED BIOSTIMULANT (ASCOPHYLLUM NODOSUM) ON ORNAMENTAL SUNFLOWER SEED GERMINATION AND SEEDLING GROWTH

The biometrics data of ornamental sunflower seedlings (Table 3) showed that height augmented as the concentration of biostimulant increased, and the best results were reached by 15 (6.9 cm) and 10 ml L⁻¹ biostimulant (5.85 cm) treatments. Plants treated with 15 mL L⁻¹ biostimulant were 54.35% higher than plants of control treatments; this result is an extremely expressive value for a program of ornamental seedlings production. Additionally, data obtained in the present study are higher than those observed by Yari et al. (2015) with 4.68 to 4.86 cm in different sunflower cultivars. Wheat seeds also had an increase of 6.7% in height when treated with seaweed extract (Kumar and Sahoo, 2011).

Table 3. Height, fresh and dry mass of shoot and root of ornamental sunflower in relation to biostimulants doses.

<table>
<thead>
<tr>
<th>Treatment (ml L⁻¹ biostimulant)</th>
<th>height cm</th>
<th>Shoot Fresh mass</th>
<th>Dry mass</th>
<th>Root Fresh mass</th>
<th>Dry mass</th>
<th>WinRhizo Length cm</th>
<th>Diameter mm</th>
</tr>
</thead>
<tbody>
<tr>
<td>0</td>
<td>3.75 c</td>
<td>0.3112 b</td>
<td>0.0150 c</td>
<td>0.085 a</td>
<td>0.0150 a</td>
<td>50.07 a</td>
<td>0.33 a</td>
</tr>
<tr>
<td>5</td>
<td>5.06 bc</td>
<td>0.3533 b</td>
<td>0.0233 bc</td>
<td>0.110 a</td>
<td>0.0144 a</td>
<td>59.56 a</td>
<td>0.34 a</td>
</tr>
<tr>
<td>10</td>
<td>5.85 ab</td>
<td>0.4480 ab</td>
<td>0.0560 b</td>
<td>0.115 a</td>
<td>0.0180 a</td>
<td>85.08 a</td>
<td>0.35 a</td>
</tr>
<tr>
<td>15</td>
<td>6.90 a</td>
<td>0.5635 a</td>
<td>0.1025 a</td>
<td>0.116 a</td>
<td>0.0195 a</td>
<td>83.65 a</td>
<td>0.34 a</td>
</tr>
<tr>
<td>DMS</td>
<td>1.65</td>
<td>0.15</td>
<td>0.04</td>
<td>0.005</td>
<td>0.009</td>
<td>54.00</td>
<td>0.11</td>
</tr>
<tr>
<td>CV (%)</td>
<td>21.98</td>
<td>25.67</td>
<td>48.77</td>
<td>36.11</td>
<td>53.00</td>
<td>29.69</td>
<td>11.96</td>
</tr>
<tr>
<td>F</td>
<td>12.95*</td>
<td>12.04*</td>
<td>23.02**</td>
<td>1.45**</td>
<td>0.84**</td>
<td>2.15**</td>
<td>10.15**</td>
</tr>
</tbody>
</table>

Means followed by the same do not differ among themselves at the 5% level of significance by the Tukey test. ns - not significant; * - significant at 5%; ** - significant at 1%.

Figure 1. Percentage of germination as a function of the time (days) of ornamental sunflower in relation to biostimulants doses.

The fresh and dry mass of shoots also augmented as the concentration of the extract increased. The concentrations 10 and 15 mL L⁻¹ biostimulant had the best results for fresh mass (p ≤ 0.5); while for dry mass the treatment 15 ml L⁻¹ biostimulant were higher (p ≤ 0.5) than other treatments (Table 3). Data related to ornamental sunflower fresh and dry mass showed higher mass (1.54-1.85 g and 1.13-1.34 g in Cabral and Castilho, 2016; 1.37-2.14 g and 0.10-0.23 g in Santos and Castilho, 2018). However, Brito et al. (2014), evaluating the fresh and dry root mass of ornamental sunflower seedlings after 15 days of germination, observed 0.2-0.7 g and 0.01-0.06 g respectively, values that are similar to those on the present study. Arabidopsis thaliana (L.) treated with A. nodosum extract increased seedling fresh mass (Rayorath et al., 2008a), probably due that seaweed extract is competent to increase genes expression responsible for endogenous auxin and cytokinin production, hormones in control of plant development (Rayorath et al., 2008b). The precise mechanism by which seaweed extracts act on plant growth and vigor has not yet been fully elucidated, but many of the effects demonstrated by the use of extracts are attributed to a variety of seaweed constituents (Wally et al., 2013), as macro and micronutrients, plant hormones analogs,
betaines and phenolic compounds (Khan et al., 2009). The application of seaweed _Ascophyllum nodosum_ extract on seeds emphasizes its physiological performance and increase seedling development (Gehling et al., 2014). Fresh and dry mass of root system and WinRhizo data had no significant statistical difference (Table 3). The root length of soybean seedlings was not affected by seaweed extract, which is probably due to the volume of tray limiting root system growth and development (Gehling et al., 2017). The size and volume of the cells in germination tray directly affects root architecture, due to restricted area that limits root growth, which may compromise shoot growth and development (Santos et al., 2019).

**Conclusions**

_Ascophyllum nodosum_ seaweed extract had a direct effect on seed germination and seedling growth variables of ornamental sunflower. Its effects were observed on the increase of seed germination variables, reduction of germination mean time and improve seedlings biomass. The concentration 15 ml L⁻¹ biosimulant ( _Ascophyllum nodosum_ extract) is recommended to enhance ornamental sunflower “Sol Pleno” seed germination and seedling growth. Additionally, this complex alga compound is natural and had no harmful effect on sunflower seeds.

**Author Contribution**

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